

## Correspondence

### Detection of a single *vanA*-containing *Enterococcus faecalis* clone in hospitals in different regions in Spain

*J Antimicrob Chemother* 2001; 48: 746–747

R. del Campo<sup>a,b\*</sup>, C. Tenorio<sup>a</sup>, M. Zarazaga<sup>a</sup>,  
R. Gomez-Lus<sup>c</sup>, F. Baquero<sup>b</sup> and C. Torres<sup>a</sup>

<sup>a</sup>Area de Bioquímica y Biología Molecular,  
Universidad de La Rioja, Logroño;

<sup>b</sup>Servicio de Microbiología, Hospital Ramón y Cajal,  
Ctra Colmenar Viejo Km 9.1, 28034 Madrid;

<sup>c</sup>Departamento de Microbiología, Universidad  
Zaragoza, Zaragoza, Spain

\*Corresponding author. Tel: +34-91-336-8330;  
Fax: +34-91-336-8809;  
E-mail: rcampo@hrc.insalud.es

Sir,

Vancomycin-resistant enterococci constitute an increasing clinical problem in the USA, and clonal dissemination of vancomycin-resistant isolates among hospitals, especially *Enterococcus faecium*, has been described.<sup>1</sup> In Europe, detection of *vanA* enterococci in the clinical setting is less frequently reported and few reports exist of clonal dissemination of resistant isolates among different hospitals.<sup>2</sup> This study examines eight *vanA*-containing *Enterococcus faecalis* clinical isolates (blood and exudates) from hospitals in four different Spanish regions: Aragón (AR721), Asturias (AS215 and AS237), Cataluña (CT715, CT716, CT718 and CT719) and La Rioja (LR337). Antibiotic resistance phenotype was determined by the agar dilution method of the NCCLS.<sup>3</sup> The putative presence of *vanR*, *vanS*, *vanH*, *vanA*, *vanY*, *vanX*, *vanZ*, *aac(6')*-*aph(2'')*, *aph(3')* and *erm(B)* genes was examined by PCR, using specific primers. Aminoglycoside-modifying enzymes were determined in extracts of resistant *Enterococcus* isolates obtained by ultrasonic disruption, using the phosphocellulose paper binding assay as described previously.<sup>4</sup> Clonal identity was studied by analysing the genomic DNA of *vanA* isolates digested with *SmaI* by pulsed-field gel electrophoresis (PFGE) as described previously.<sup>5</sup> Isolates were classified as indistinguishable, closely related, possibly related or unrelated according to published criteria.<sup>6</sup>

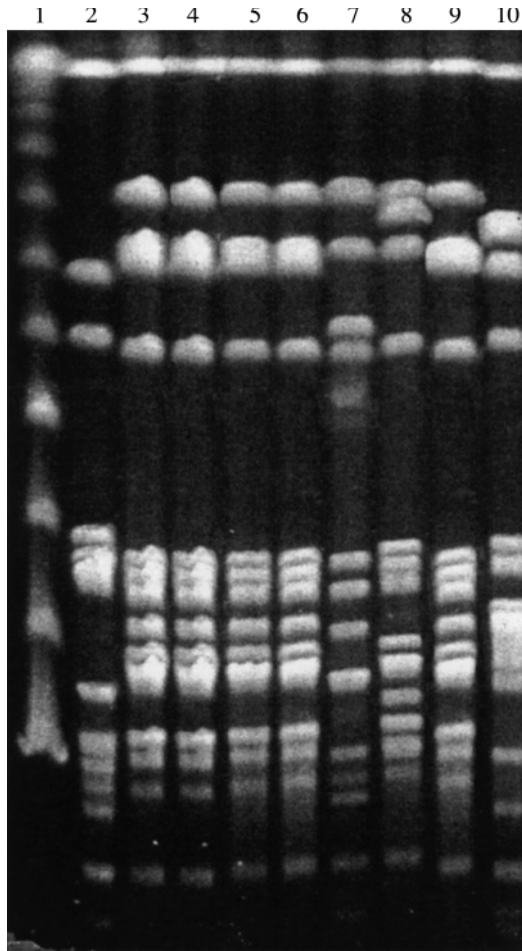
All eight *vanA* *E. faecalis* isolates were resistant to vancomycin and teicoplanin (MIC ranges 256–512 mg/L and 64–256 mg/L, respectively) as well as to erythromycin

(MIC > 128 mg/L), but were susceptible to ampicillin (MIC < 4 mg/L). Seven isolates showed high-level resistance (HLR) to kanamycin (>2000 mg/L) and streptomycin (>1000 mg/L); one (AS237) also showed HLR to gentamicin (>1000 mg/L) and one (CT718) showed HLR only to streptomycin. In *E. faecalis* strains that showed HLR to kanamycin APH(3') activity was detected by the radioenzymic assay and the presence of the *aph3'*-III gene was confirmed by PCR amplification. No streptomycin-modifying enzyme was detected in high-level streptomycin-resistant isolates. APH(2'')-AAC(6') activity was detected in the *E. faecalis* strain that showed HLR to gentamicin, and the presence of the *aac6-aph2* gene was confirmed by PCR. Positive PCR amplifications were obtained in all eight isolates for all genes of the *vanA* operon (*vanR*, *vanS*, *vanH*, *vanA*, *vanX*, *vanY* and *vanZ*) and also for the *erm(B)* gene.

All *E. faecalis* isolates carried a high molecular weight plasmid (c. 60 kb) that was transferred to the recipient *E. faecalis* JH2-2 strain by filter mating. Both donors and transconjugants also showed a positive hybridization pattern with a *vanA* probe in the chromosome. The same hybridization pattern was obtained when genomic DNA was digested with *EcoRI*, transferred to a nylon membrane and hybridized with the *vanA* probe.

Five unrelated patterns were found by *SmaI*-PFGE among the eight *E. faecalis* isolates. Four *vanA*-carrying *E. faecalis* isolates from three geographically distant hospitals showed an indistinguishable PFGE pattern (*E. faecalis* CT716, CT719, LR337 and AS215) (Figure). These four isolates were also further characterized as bacteriocin producers with a broad spectrum of activity,<sup>7</sup> similar in all four strains. The other four *E. faecalis* isolates (CT715, CT718, AR721 and AS237) showed unrelated PFGE patterns; from these, one strain was a non-bacteriocin producer, and the other three produced possibly different antibacterial substances.

The genetic data shown in this work indicate that a group of four *E. faecalis* isolates (CT716, CT719, LR337 and AS215) recovered from clinical samples in hospitals from three geographically distant Spanish regions constitute a single clone. To our knowledge, this is the first time that a single clone of *vanA*-carrying *E. faecalis* has been detected in different geographically separate hospitals. An alternative explanation for this event could be the dissemination of a particular vancomycin-susceptible *E. faecalis* clone among the three Spanish regions that may have independently acquired the *vanA* operon, erythromycin and aminoglycoside resistance genes. Nevertheless, the unique spectrum of bacteriocin activity of these *E. faecalis* isolates<sup>7</sup>



**Figure.** *Sma*I-PFGE of eight *E. faecalis vanA* isolates from various hospitals in separate regions of Spain. Lane 1: PFGE marker; lanes 2–10: *E. faecalis vanA* AS237, AS215, CT716, CT719, LR337, CT715, CT718, AR721 and LRH1.

suggests a single event of *vanA* acquisition, followed by clonal dissemination in different regions.

## Acknowledgements

We are grateful to M. Lantero, J. Castillo, C. Rubio, F. Marco and A. Fleites for submitting the *E. faecalis vanA* isolates. R.d.C. was supported by a grant from the Diputación General de Aragón (project P94/97). This work has been supported in part by a grant from the Fondo de Investigaciones Sanitarias (00/0545), Spain.

## References

1. Chow, J. W., Kuritzin, A., Shalae, D. M., Green, M., Sahm, D. F. & Zervos, M. J. (1993). Clonal spread of vancomycin-resistant *Enterococcus faecium* between patients in three hospitals in two states. *Journal of Clinical Microbiology* **31**, 1609–11.
2. Nourse, C., Byrne, C., Kaufmann, M., Keane, C. T., Fenelon, L., Smyth, E. G. *et al.* (2000). VRE in the Republic of Ireland: clinical significance, characteristics and molecular similarity of isolates. *Journal of Hospital Infection* **44**, 288–93.
3. National Committee for Clinical Laboratory Standards. (2001). *Methods for Dilution Antimicrobial Susceptibility Testing for Bacteria that Grow Aerobically: Approved Standard M7-A5*. NCCLS, Wayne, PA.
4. Haas, M. J. & Dowding, J. E. (1975). Aminoglycoside-modifying-enzymes. *Methods in Enzymology* **43**, 611–40.
5. Murray, B. E., Singh, K. V., Heath, J. D., Sharma, B. R. & Weinstein, G. M. (1990). Comparison of genomic DNAs of different enterococcal isolates using restriction endonucleases with infrequent recognition sites. *Journal of Clinical Microbiology* **28**, 2059–63.
6. Tenover, F. C., Arbeit, R. D., Goering, R. V., Mickelsen, P. A., Murray, B. E., Persing, D. H. *et al.* (1995). Interpreting chromosomal DNA restriction patterns produced by pulsed field gel electrophoresis: criteria for bacterial strain typing. *Journal of Clinical Microbiology* **33**, 2233–9.
7. Del Campo, R., Tenorio, C., Jimenez-Diaz, R., Rubio, C., Gomez-Lus, R., Baquero, F. *et al.* (2001). Bacteriocin production in vancomycin-resistant and vancomycin-susceptible *Enterococcus* isolates of different origins. *Antimicrobial Agents and Chemotherapy* **45**, 905–12.